KIDNEY FAILURE

Carbamylation of Serum Albumin as a Risk Factor for Mortality in Patients with Kidney Failure

Anders H. Berg,¹* Christiane Drechsler,² Julia Wenger,³ Roberto Buccafusca,^{4,5} Tammy Hod,⁴ Sahir Kalim,³ Wenda Ramma,¹ Samir M. Parikh,⁴ Hanno Steen,⁵ David J. Friedman,⁴ John Danziger,⁴ Christoph Wanner,² Ravi Thadhani,³ S. Ananth Karumanchi^{4,6}

Urea, the toxic end product of protein catabolism, is elevated in end-stage renal disease (ESRD), although it is unclear whether or how it contributes to disease. Urea can promote the carbamylation of proteins on multiple lysine side chains, including human albumin, which has a predominant carbamylation site on Lys⁵⁴⁹. The proportion of serum albumin carbamylated on Lys⁵⁴⁹ (%C-Alb) correlated with time-averaged blood urea concentrations and was twice as high in ESRD patients than in non-uremic subjects (0.90% versus 0.42%). Baseline %C-Alb was higher in ESRD subjects who died within 1 year than in those who survived longer than 1 year (1.01% versus 0.77%) and was associated with an increased risk of death within 1 year (hazard ratio, 3.76). These findings were validated in an independent cohort of diabetic ESRD subjects (hazard ratio, 3.73). Decreased concentrations of serum amino acids correlated with higher %C-Alb in ESRD patients, and mice with diet-induced amino acid deficiencies exhibited greater susceptibility to albumin carbamylation than did chow-fed mice. In vitro studies showed that amino acids such as cysteine, histidine, arginine, and lysine, as well as other nucleophiles such as taurine, inhibited cyanate-induced C-Alb formation at physiologic pH and temperature. Together, these results suggest that chronically elevated urea promotes carbamylation of proteins in ESRD and that serum amino acid concentrations may modulate this protein modification. In summary, we have identified serum %C-Alb as a risk factor for mortality in patients with ESRD and propose that this risk factor may be modifiable with supplemental amino acid therapy.

INTRODUCTION

Chronic kidney disease (CKD) affects 5 to 10% of adults in industrialized countries (1). For reasons that remain unclear, individuals with CKD are 10 to 20 times more likely to die from cardiovascular causes than to survive until renal function is completely lost (2). Those who reach end-stage renal disease (ESRD) suffer an annual mortality of 15 to 20% that is largely attributable to cardiovascular disease (CVD) (3). Yet, efforts to treat the most modifiable cardiovascular risk factor, hypercholesterolemia, with statins have not improved outcomes in ESRD (4, 5). This finding suggests that other mechanisms link ESRD to CVD (6).

One possible mechanism for how ESRD increases the risk of CVD is the accumulation of urea in the blood of ESRD patients. Urea is generated in the liver during catabolism of amino acids and other nitrogenous metabolites and is normally excreted into the urine by the kidneys as rapidly as it is produced. Patients with ESRD cannot make urine, however, and thus, increasing concentrations of blood urea will steadily accumulate, a condition that can only be treated with intermittent hemodialysis (HD) or kidney transplantation. Although HD ameliorates ESRD patients' uremia, it replaces only ~10% of normal renal function, so these patients still have chronic urea overload (uremia).

Despite the strong association between ESRD and CVD, the role of chronically elevated urea in this disease is controversial. For example, average urea concentration does not predict mortality in CKD, and

the HEMO study found no benefit for survival when frequency of HD was increased beyond the current standards (7-10). This lack of effect could have been because the ~40% relative increase in waste removal only replaces ~14% of normal kidney filtration and still leaves patients overloaded with waste products. Recent evidence suggests that chronically elevated blood urea contributes directly to cardiovascular risk via a proatherogenic protein modification called carbamylation. In one study of patients undergoing diagnostic cardiac catheterization, subjects in the highest quartile of serum protein-bound carbamylated lysine had a seven to eight times higher risk of CVD; these authors also demonstrated that low-density lipoprotein (LDL) was a target for protein carbamylation and that carbamylated LDL (cLDL) binds scavenger receptors and produces lipid accumulation in macrophages (11). In addition, feeding urea to apolipoprotein E (apoE)-deficient mice accelerated their rate of atherosclerosis nearly twofold and increased accumulation of cLDL within atherosclerotic plaques (12). Together, these findings suggest a potential mechanism for urea's direct contribution to atherogenesis.

Protein carbamylation is an unavoidable consequence of excess urea (13, 14). Urea is in equilibrium with cyanate (HNCO), a product of urea deamination whose central carbon is susceptible to nucleophilic attack from amines at the N termini or side chains of proteins in vivo (15). The degree to which proteins with long half-lives are carbamylated should therefore provide a time-averaged indicator of urea concentration, analogous to the relationship between serum glucose and glycated hemoglobin (16). Thus, measurements of carbamylated proteins in circulation might provide a quantitative biomarker that is mechanistically linked to urea kinetics and the effectiveness of urea reduction therapy. If carbamylated proteins are indicators of chronic uremia and also direct contributors to atherogenesis and cardiovascular pathology (11, 12), then concentrations of circulating carbamylated proteins should be closely correlated with ESRD patients' cardiovascular health and overall mortality.

¹Division of Clinical Chemistry, Department of Pathology, Beth Israel Deaconess Medical Center, Boston, MA 02215, USA. ²Division of Nephrology, Department of Internal Medicine, University of Würzburg, Würzburg D-97074, Germany. ³Division of Nephrology, Department of Medicine, Massachusetts General Hospital and Harvard Medical School, Boston, MA 02114, USA. ⁴Division of Nephrology and Center for Vascular Biology Research, Department of Medicine, Beth Israel Deaconess Medical Center and Harvard Medical School, Boston, MA 0215, USA. ⁵Department of Pathology, Boston Children's Hospital and Harvard Medical School, Boston, MA 0215, USA. ⁵To whom correspondence should be addressed. E-mail: ahberg@bidmc.harvard.edu

We therefore measured carbamylation of the long-lived circulating protein albumin and tested whether albumin carbamylation correlated with outcomes in two independent cohorts of patients with ESRD. Furthermore, we tested whether higher-protein carbamylation is associated with increased average blood urea concentrations and whether protein carbamylation can be competitively inhibited by free amino acids.

RESULTS

A preferred carbamylation site on albumin and validation of a high-throughput assay

To map sites on circulating albumin susceptible to carbamylation, cyanate-treated albumin was digested with proteinase, digested peptides were separated by high-performance liquid chromatography (HPLC), and peptides were analyzed for carbamylation modifications with tandem mass spectrometry (MS/MS). Several carbamylation modifications were detected, the most abundant on Lys⁵⁴⁹ (Fig. 1). The relative amount of carbamylation at each site was quantified by measuring its carbamylated and noncarbamylated peptide forms with multiple reaction monitoring (MRM) and then calculating their ratio (table S1). The susceptibility to carbamylation at Lys⁵⁴⁹ is consistent with reports that Lys⁵⁴⁹ is also a common target for glycation (17). We then developed a high-throughput assay that incorporated isotopic peptide standards to reproducibly quantitate the proportion of albumin carbamylated on Lys⁵⁴⁹ (%C-Alb) as described in Materials and Methods (fig. S1). Using this assay, we demonstrated that average %C-Alb was about twofold higher in patients with CKD and ESRD than in non-uremic subjects

Protein Pilot MS/MS peptide identification summary report



*Predicted ions found in spectra are shown in red

Fig. 1. Identification of carbamylation on Lys⁵⁴⁹ of cyanate-treated albumin. Cyanate-treated and untreated purified human albumin from a commercial source was digested with glutamyl endoproteinase and analyzed by liquid chromatography–MS/MS (LC-MS/MS). Peptides were identified by matching fragmentation spectra to sequences from the human Swiss-Prot proteome with the AB Sciex Protein Pilot software programmed to search for both carbamylated and noncarbamylated peptide forms. (**A**) Summary reports for the carbamylated and noncarbamylated forms of the peptide encompassing Lys⁵⁴⁹ (sequence **RQIKXQTALVE**, where **X** = *N*- ε -carbamoyl-L-lysine). Contribution of identified peptide (in ProtScore units) to protein identification; Conf, percent confi-

dence of peptide identification; *z*, peptide charge; Spectrum, within-run identifier for MS/MS spectra used for peptide identification. (**B**) MS/MS spectra for carbamylated and noncarbamylated digested albumin peptides encompassing Lys⁵⁴⁹; the modification state–revealing fragment ions are shown in red. (**C**) Predicted ion fragments of carbamylated and noncarbamylated peptide forms; detected fragment ions matching the predicted fragmentation spectra are highlighted in red. Both peptides were identified with 99+% confidence using the Paragon algorithm, which compares the confidence of the match between the observed spectra and the identified peptide sequence to the combined confidence for all other possible peptide matches (*54*).

(Fig. 2). Furthermore, %C-Alb values were highly correlated with the proportional amounts of carbamylated lysine in the unfractionated serum proteins of uremic patients and were correlated with amounts of percent carbamylated hemoglobin in these patients (fig. S2). These correlations corroborate the association between %C-Alb and global carbamylation of blood proteins in uremic subjects.

Thus, Lys⁵⁴⁹ is the most frequently carbamylated amino acid on human albumin, although there are additional sites that are less susceptible to carbamylation. Furthermore, we developed a high-throughput assay for measuring carbamylated albumin that can be used in large clinical studies and confirmed that %C-Alb values were elevated in patients with uremia.

Association of %C-Alb with mortality in patients on HD

%C-Alb was measured in serum from 187 participants from the Accelerated Mortality on Renal Replacement (ArMORR) study (*18*, *19*). Subjects included 81 ArMORR participants who died from any cause during 12 months of follow-up (ArMORR cases) and 106 specimens from ArMORR subjects who survived longer than 365 days (ArMORR survivors). The 12-month follow-up period began 90 days after initiation of HD. Subjects' baseline clinical characteristics and biochemical analyses were measured on day 90 of the study run-in period and are presented in table S3. As shown in Fig. 2, ArMORR cases who died during follow-up had significantly higher average baseline %C-Alb values than did ArMORR survivors (1.01% versus 0.77%, *P* = 0.0009; Fig. 2). Nevertheless, the concentration of urea in their blood was not different from that of ArMORR survivors (16.9 mM versus 17.3 mM, respectively, *P* = 0.72), suggesting that factors other than urea concentration may also regulate %C-Alb values.

We analyzed %C-Alb values for their association with 12-month mortality by using Kaplan-Meier survival analysis (20), which showed statistically significant higher mortality for ArMORR subjects in the



Fig. 2. Average carbamylated albumin values in uremic and non-uremic patients. (**A**) Average %C-Alb values in non-uremic subjects (n = 20) and in patients with stage 3 or 4 CKD (n = 122) and ArMORR HD subjects (n = 187). (**B**) Average %C-Alb in ArMORR survivors who lived longer than 12 months (n = 106) and in ArMORR cases who died during the 12-month study period (n = 81). Individual %C-Alb values for each group are shown in table S2. Data are expressed as average carbamylated albumin as a percentage of total; error bars, 95% confidence intervals (Cls) of the mean; Student's *t* test *P* values are shown.

highest %C-Alb tertile than for subjects with less %C-Alb (Fig. 3). Using a Cox proportional hazards model (21), we found that increased %C-Alb values were associated with significantly increased risk of death [hazard ratio (HR), 3.76; 95% CI, 2.20 to 6.43; P < 0.0001] (Table 1). Statistically significant risks of death were also associated with decreased albumin, decreased hemoglobin, and a history of hypertension, which are all known risk factors for death in patients with ESRD. Nevertheless, the risk of death associated with %C-Alb remained strong and statistically significant even after adjusting for these other risk factors (HR, 3.23; 95% CI, 1.74 to 6.00; P = 0.0002). There was no significant risk associated with the subjects' baseline equilibrated Kt/V values, a measure of urea reduction and dialysis adequacy (22). Furthermore, when we excluded ArMORR survivors from the model and stratified just the ArMORR cases by their %C-Alb values, we observed that the highest %C-Alb tertile within this subgroup was associated with an increased hazard rate compared to the cases in the bottom two tertiles (univariate HR, 1.56; 95% CI, 1.00 to 2.43; P =0.04). This indicated that even among the cohort ArMORR subjects that died during the 12-month study, there was an association between higher %C-Alb values and shorter survival times.

To validate the major findings from the ArMORR cohort, we tested whether baseline %C-Alb was associated with mortality in a second independent study of HD subjects. We obtained frozen specimens from 1161 ESRD patients who had participated in the 4D (Deutsche Diabetes Dialyze Studie) HD trial, a randomized controlled trial of the benefits of cholesterol-lowering agents in HD patients with diabetes mellitus (4, 23). Baseline characteristics of 4D subjects are shown in table S4. Similar to the ArMORR study, %C-Alb was measured in specimens sampled at the beginning of the study and analyzed for associations to survival during a 12-month follow-up period. Kaplan-Meier survival analysis and multivariable HR analysis of %C-Alb and Kaplan-Meier survival analysis again found significant risk of death among 4D subjects with elevated %C-Alb (Table 2 and Fig. 3), with HR estimates similar to those observed in the ArMORR study. When we analyzed the risk of death in ArMORR and 4D subjects stratified into tertiles according to their %C-Alb values, there was again a gradation of risk between subjects stratified by their %C-Alb values (table S5).

Kidney disease is frequently associated with diabetes mellitus; it is thus possible that risks associated with diabetes mellitus may confound the association between %C-Alb and risk of death. Comparison of the univariate and multivariate HRs associated with %C-Alb showed that adjustment for ArMORR subjects' history of diabetes mellitus or 4D subjects' duration of diabetes mellitus did not significantly change the risk associated with %C-Alb for either of these studies (Tables 1 and 2), however. In summary, data from two large independent clinical trials suggested that %C-Alb may be an independent risk factor for mortality in ESRD patients.

%C-Alb as an index of urea overload and circulating free amino acids

To test whether nucleophilic amines and thiols of free amino acids compete with the side chains of albumin for carbamylation, we measured free amino acids and their carbamylated forms in ArMORR study subjects and analyzed their %C-Alb values as a composite function of serum-free amino acid and urea concentrations. We observed a modest positive correlation between time-averaged urea and %C-Alb in ArMORR subjects (Fig. 4A) and a similar correlation between urea and %C-Alb in 4D study subjects (Fig. 4B). The correlation between



Fig. 3. Kaplan-Meier curve estimates of the incidence of all-cause mortality in ESRD patients. Subjects were categorized into upper, middle, and lower tertiles according to serum %C-Alb values measured at the outset of the study. (A)

Twelve-month survival rates in ArMORR study subjects. (**B**) Twelve-month survival rates in 4D study subjects. Numbers of subjects at risk at different time points during each study are shown in the tables at the bottom.

urea and %C-Alb was significantly stronger in patients with stage 3 or 4 CKD than in either ESRD cohort (Fig. 4C).

In contrast to the positive correlation between blood urea concentrations and %C-Alb, individual serum-free amino acid concentrations in our ArMORR subjects were negatively correlated with %C-Alb, suggesting an association between low free amino acid concentrations and high serum protein carbamylation (Table 3). To test whether free amino acid levels could influence the correlation between uremia and higher %C-Alb, we examined these relationships while adjusting for the other variables. In most cases, we found stronger correlations between %C-Alb and urea after adjusting for amino acid concentrations (Table 3). Together, these studies suggest that increased urea and decreased amino acid concentrations are independently correlated with protein carbamylation in ESRD patients.

Susceptibility of different amino acids to carbamylation in vitro

We postulated that serum amino acids may correlate with %C-Alb because they are alternative substrates for carbamylation by cyanate. To differentiate the cyanate-scavenging ability of individual amino acids, we added cyanate to an equimolar mixture of all 20 amino acids. As expected, increasing concentration of cyanate produced linear increases in all carbamylated amino acids (fig. S3A) and depletion of their unmodified forms (fig. S3, B and C). The percent depletion for each amino acid in the presence of 5 mM cyanate is a quantitative index of their relative avidities within this competitive carbamylation reaction (fig. S3D). The dipeptide glycylglycine was more strongly carbamylated than glycine or other amino acids. These experiments demonstrated the comparative susceptibilities and competition among individual amino acids for carbamylation and suggested that amino acids may compete with proteins for carbamylation as well.

Prevention of albumin carbamylation by amino acids and other nucleophiles

To test whether competing nucleophiles can prevent albumin carbamylation by cyanate, we then tested whether individual amino acids can reverse carbamylation of purified serum albumin. Although all amino acids that we tested were able to inhibit carbamylation, some amino acids (cysteine, histidine, and arginine) were more effective **Table 1.** In the ArMORR study, HR estimates for 12-month mortality (risk of death from all causes over the 12 months after the 90-day study runin period) associated with %C-Alb and other parameters. Values were measured at baseline (start of the study). Additional patient characteristics were included in this analysis but demonstrated no statistically significant risk in univariate analysis. These variables included body mass index; leukocyte count; platelet count; serum concentrations of potassium, phosphorus, calcium, ferritin, alkaline phosphatase, and parathyroid hormone; and percentage of subjects with histories of diabetes mellitus, coronary artery disease/myocardial infarction, chronic obstructive pulmonary disease, cancer, dyslipidemia, anemia, peripheral vascular disease, cerebrovascular accident, congestive heart failure, atrial fibrillation, and liver disease.

Variables	Univariate		Multivariate	
	HR (95% CI)	Р	HR (95% CI)	Р
%C-Alb*	3.76 (2.20–6.43)	< 0.0001 ⁺	3.23 (1.74–6.00)	0.0002 [†]
Albumin	0.43 (0.26-0.71)	0.001 ⁺	0.53 (0.30–0.93)	0.03 [†]
Hemoglobin	0.79 (0.69–0.91)	0.001 ⁺	0.96 (0.82–1.14)	0.65
History of hypertension	0.42 (0.25-0.71)	0.001 ⁺	0.42 (0.23–0.74)	0.003 ⁺
Equilibrated Kt/V	0.74 (0.30–1.80)	0.50	0.95 (0.37–2.41)	0.91
Systolic blood pressure	0.98 (0.97–0.99)	< 0.0001 ⁺	0.99 (0.98–1.00)	0.05
History of diabetes mellitus	0.69 (0.40–1.17)	0.17	1.03 (0.55–1.91)	0.94

*Continuous variable, natural log-transformed. *P* values considered significant at *P* < 0.05.

Table 2. In the 4D study, HR estimates for 12-month mortality (risk of death from all causes over the 12 months after the 90-day study run-in period) associated with %C-Alb and other parameters. Values were measured at baseline (start of the study). Additional patient characteristics were also included in this analysis but demonstrated no statistically signif-

inhibitors than others (Table 4). The dipeptide glycylglycine reduced the amount of albumin carbamylation by 63%. Taurine, the most abundant intracellular amine, also effectively scavenged cyanate. These results demonstrated that carbamylation of albumin can be counteracted by free amino acids and other endogenous nucleophilic biomolecules.

Protein carbamylation in mice with amino acid deficiencies

To further test our hypothesis that amino acid deficiencies cause animals to be more susceptible to protein carbamylation, we induced amino acid deficiencies by feeding mice low-protein diets for 15 days. Mice were also treated with either cyanate or urea to induce protein carbamylation. In the first experiment, we tested the effect of a low-protein diet on acute protein carbamylation: On day 15, animals were injected with cyanate; 30 min after injection, blood was drawn and tested for serum %C-Alb and free amino acid concentrations. Animals on the low-protein diet displayed significantly lower serum essential amino acid concentrations than did control animals fed a normal-protein diet (Fig. 5A). When the amino acid–deficient mice were then injected with cyanate, they showed twofold greater increases in %C-Alb values than did the animals on the normal-protein diet (Fig. 5B).

In the second mouse experiment, the effects of chronically elevated blood urea were tested by adding urea to the animals' food. Four groups of mice were fed for 15 days with low- or normal-protein diets mixed with urea in doses roughly equivalent to the animals' normal daily urea output (24). On day 15, blood was drawn and serum was

icant differences between cases and controls. These variables included gender; systolic and diastolic blood pressure; platelet count; serum concentrations of parathyroid hormone, potassium, calcium, phosphorus, ferritin, and LDL cholesterol; and percentage of subjects with histories of diabetes mellitus, hypertension, or cerebrovascular accident.

Variables	Univariate	Univariate		Multivariate	
	HR (95% CI)	Р	HR (95% CI)	Р	
%C-Alb*	3.73 (2.00–6.96)	<0.001 ⁺	3.17 (1.55–6.46)	0.002 ⁺	
Age	1.04 (1.02–1.06)	<0.001 ⁺	1.03 (1.00–1.05)	0.050	
Body mass index	0.94 (0.91–0.97)	0.001 ⁺	0.97 (0.93–1.01)	0.122	
Alkaline phosphatase	1.004 (1.002–1.005)	<0.001 ⁺	1.002 (1.001–1.004)	0.008 ⁺	
Albumin	0.50 (0.30–0.82)	0.006 ⁺	0.90 (0.50–1.63)	0.723	
Hemoglobin	0.87 (0.77–0.97)	0.017 ⁺	0.94 (0.82–1.08)	0.366	
Transferrin saturation	0.98 (0.97-1.00)	0.016 ⁺	0.99 (0.97–1.01)	0.180	
Leukocyte count	1.12 (1.06–1.19)	<0.001 ⁺	1.09 (1.02–1.16)	0.007 ⁺	
Coronary artery disease	1.54 (1.11–2.13)	0.009 ⁺	1.22 (0.83–1.77)	0.312	
Congestive heart failure	1.61 (1.18–2.21)	0.003 ⁺	1.20 (0.83–1.75)	0.338	
Peripheral vascular disease	2.01 (1.46–2.78)	<0.001 ⁺	1.82 (1.25–2.66)	0.002 ⁺	
Atrial fibrillation	2.24 (1.48–3.39)	<0.001 ⁺	1.57 (0.92–2.66)	0.097	
Duration of diabetes (years)	1.03 (1.01–1.05)	0.003 ⁺	1.03 (1.01–1.05)	0.010 ⁺	
Urea reduction ratio	1.00 (0.97–1.02)	0.857			
% Hemoglobin A _{1c}	1.05 (0.93–1.19)	0.408			

*Continuous variable transformed to its natural log. \uparrow values considered significant at P < 0.05.



Fig. 4. Correlation between %C-Alb and blood urea concentrations. (**A**) Correlation between blood urea and %C-Alb in ArMORR study subjects with ESRD (n = 187). (**B**) Correlation between blood urea and %C-Alb in 4D study subjects with ESRD (n = 1161). (**C**) Correlation between %C-Alb and blood urea in non-HD CKD subjects (n = 122). Pearson correlation coefficients (r) and P values are shown.

analyzed for %C-Alb and free amino acids. The amino acid-deficient mice fed the low-protein diet along with urea had significantly increased %C-Alb compared to urea-fed animals on the normal-protein diet Fig. 5C. In summary, mouse models of both acute and chronic albumin carbamylation confirmed that animals with amino acid deficiencies experience markedly increased degrees of carbamylation in the presence of equivalent doses of cyanate or urea.

Potential confounders of %C-Alb measurements

Kidney disease is frequently associated with diabetes mellitus and hyperglycemia. Lys⁵⁴⁹ on albumin is susceptible to both carbamylation and glycation by glucose (17). To evaluate whether glycated albumin interferes with accurate assessment of %C-Alb values, we modified our %C-Alb assay to also measure glycation of Lys⁵⁴⁹. We analyzed %C-Alb and percent glycated albumin in 40 serum samples from patients with both diabetes mellitus and chronic or end-stage renal disease. We observed no statistically significant correlation between carbamylated and glycated albumin values, however, suggesting that glycation at Lys⁵⁴⁹ does not interfere with the carbamylation reaction at that same site (fig. S4). Furthermore, multivariable-adjusted HR analysis demonstrated that the association between %C-Alb and mortality was independent of history or duration of diabetes mellitus (Tables 1 and 2).

Calculation of %C-Alb includes the measured concentration of noncarbamylated albumin in the denominator. Low serum albumin concentration (hypoalbuminemia) is a known risk factor in patients with renal disease (25), and so, we considered whether the risk associated with %C-Alb was confounded by the effects of low serum albumin concentrations. ArMORR subjects' %C-Alb values and serum albumin concentrations showed no significant correlation to each other, however (fig. S5).

In summary, evaluation of important risk factors associated with ESRD (diabetes mellitus and hypoalbuminemia) showed no correlations between these variables and %C-Alb values and no confounding effects by these variables on the association between %C-Alb and risk of death.

DISCUSSION

We hypothesized that serum protein carbamylation reflects a balance between urea load from kidney filtration failure and amino acid deficiency arising from multiple factors, including the undernourishment common to CKD (26). To study this, we developed an assay to quantify the carbamylation of serum albumin in vivo and applied this method to subjects with ESRD, a condition of chronic urea overload. We analyzed samples from two previously published large independent cohorts and found that %C-Alb was strongly associated with 1-year mortality in patients with ESRD undergoing HD. Furthermore, the association between %C-Alb and 1-year mortality was undiminished even after adjustment for other significant risk factors for mortality (age, body mass index, history of coronary artery disease, congestive heart disease, peripheral vascular disease, atrial fibrillation, hypertension, systolic blood pressure, history and duration of diabetes mellitus, blood hemoglobin concentration, serum albumin, transferrin saturation, and blood leukocyte counts). In contrast, we found that standard measures of dialysis adequacy (equilibrated Kt/V and urea reduction ratio) were not associated with risk of death. We also examined the effect of free amino acids on %C-Alb in vitro. In studies in human subjects, amino acid-deficient mice, and in vitro experiments, we have gathered evidence in support of the conclusion that low free amino acid concentrations and high urea load combine to favor formation of carbamylated albumin. We have identified select amino acids (cysteine and histidine) and similar nucleophilic biomolecules (glycylglycine, taurine, and cysteamine) that can act as potent inhibitors of protein carbamylation. Together, these data identify **Table 3.** Spearman correlations and partial correlations between %C-Alb, blood urea, and free amino acids in ArMORR study subjects.

Amino acids	Unadjusted correlation*		Partial correlation to urea, adjusted for individual amino acids [†]	
	rs	Р	Partial r _s	Partial P
Average urea	0.431	<0.0001 [‡]		
Arginine	-0.357	0.0004 [‡]	0.507	<0.0001 [‡]
Lysine	-0.310	0.0022 [‡]	0.454	<0.0001 [‡]
Histidine	-0.270	0.0082 [‡]	0.477	<0.0001 [‡]
Alanine	-0.341	0.0007 [‡]	0.434	<0.0001 [‡]
Glycine	-0.216	0.0354 [‡]	0.406	<0.0001 [‡]
Threonine	-0.105	0.3121	0.430	<0.0001 [‡]
Serine	-0.270	0.0081 [‡]	0.464	<0.0001 [‡]
Proline	-0.174	0.0914	0.425	<0.0001 [‡]
Glutamine	-0.238	0.0204 [‡]	0.474	<0.0001 [‡]
Methionine	-0.094	0.3672	0.423	<0.0001 [‡]
Tyrosine	-0.080	0.4384	0.433	<0.0001 [‡]
Valine	-0.240	0.0193 [‡]	0.488	<0.0001 [‡]
Leucine/isoleucine	-0.388	0.0001 [‡]	0.484	<0.0001 [‡]
Aspartic acid	-0.218	0.0338 [‡]	0.450	<0.0001 [‡]
Glutamic acid	-0.060	0.5629	0.429	<0.0001 [‡]
Phenylalanine	-0.134	0.1939	0.458	<0.0001 [‡]
Tryptophan	-0.100	0.3352	0.423	<0.0001 [‡]

*Unadjusted Spearman rank correlations compared to %C-Alb. †Partial Spearman correlations between average predialysis urea values and %C-Alb after adjustment for amino acid concentrations measured at the start of the study. P values considered significant at P < 0.05. r_s indicates Spearman correlation coefficients with %C-Alb.

%C-Alb as a possible prognostic indicator for adverse outcomes in ESRD and suggest that the levels of this indicator may be modified by free amino acid supplementation in patients.

Carbamylation of free amino acids and proteins by urea-derived cyanate was first described in 1960 (15), has been studied primarily in kidney failure and uremia (11, 12, 14, 27-40), and was reviewed recently in (41). Whether excess urea is pathogenic has been a controversial issue. Medical textbooks often assert that urea is relatively nontoxic, but the evidence most often cited for this claim is a 40year-old pilot trial of three dialysis subjects who were loaded with excess urea over a period of 90 days and monitored for acute symptoms of uremia. Although the acute symptoms of urea loading were relatively minor, the study did not include any follow-up for longterm effects (7). Opinions regarding urea's toxicity by the medical community were also influenced by the HEMO clinical trial, in which it was found that increasing HD dose (as indicated by equilibrated Kt/V) was ineffective for improving survival in patients (10, 42). The results from HEMO may be uninformative because more frequent dialysis had a relatively small effect on urea removal, however, so it is not surprising that no survival benefit could be detected (43). More recent studies testing the effects of increased frequency of **Table 4.** Inhibition of in vitro carbamylation by amino acid scavengers. Purified albumin was premixed with or without individual amino acids (10 mM) and carbamylated overnight at 37° C with 0.5 mM cyanate and then analyzed for %C-Alb.

Amino acid	%C-Alb (mean ± SD)	P *	% Decrease compared to no inhibitor [†]
Glycylglycine	15 ± 1.2%	<0.0001	64
Cysteine + cysteamine	17 ± 0.9%	<0.0001	59
Taurine	22 ± 1.7%	<0.0001	46
Cysteamine	23 ± 1.0%	<0.0001	42
Cysteine	23 ± 1.9%	<0.0001	42
Histidine	26 ± 1.0%	<0.0001	35
Arginine	27 ± 1.6%	<0.0001	34
Glutathione	29 ± 1.4%	<0.0001	28
Lysine	31 ± 0.9%	0.0002	23
Glycine	31 ± 1.8%	0.006	23
Glutamine	32 ± 2.9%	0.009	21
Tryptophan	32 ± 2.7%	0.012	21
Alanine	32 ± 1.7%	0.018	20
Valine	33 ± 1.1%	0.001	18
Proline	34 ± 1.6%	0.006	16
Leucine	37 ± 2.2%	0.035	10
Glutamate	38 ± 1.2%	0.20	6
No amino acid*	41 ± 2.5%*		
No cyanate	0.17 ± 0.2%		

**P* values indicate significance of differences between the average %C-Alb values in samples premixed with amino acid inhibitors compared to no amino acid added. Percent decrease in carbamylation was calculated by taking the difference between %C-Alb in samples with no amino acid added (41%) and samples with amino acid inhibitors added and divided by 41%. For example, the percent decrease in carbamylation for alanine is (41% - 32%)/41% = 20% inhibition of protein carbamylation.

HD—comparing daily dialysis to the standard prescription of three times per week—did show benefits for the subjects (44, 45). Thus, it now appears that the toxicity of urea may have been dismissed prematurely.

The most common cause of death in our study subjects (and in ESRD patients in general) was a cardiac event, and a growing body of evidence has demonstrated that protein carbamylation is associated with risk of CVD in patients. Patients undergoing cardiac catheterization who had increased serum carbamylated proteins were at significantly increased risk of CVD, coronary heart disease, and peripheral vascular disease (11), and higher carbamylated lysine content in serum proteins-a marker of protein carbamylation-was significantly associated with 5-year mortality in uremic patients on HD (46). Studies from tissue culture and animal models also suggest that urea and protein carbamylation may contribute to CVD. First, carbamylated albumin appears to be proinflammatory (47). Second, carbamylated proteins are highly enriched in atherosclerotic plaques, and cLDL may be a pathogenic ligand for foam cells (11, 28). In addition to transporting cholesterol into atherosclerotic plaques, cLDL also induces endothelial cell death, vascular smooth muscle cell proliferation, monocyte inflammatory



Fig. 5. Effects in mice of low-protein diet on albumin carbamylation by urea or cyanate. (**A**) Serum amino acid concentrations after 15 days of low-or normal-protein diets. Values are normalized to average concentrations of amino acid in animals on a normal-protein diet. **P* < 0.05 with an unpaired *t* test, comparing average amino acids in animals on low- and normal-protein diets. (**B**) %C-Alb values in animals fed low- or normal-protein diets before and 30 min after cyanate injection (100 mg/kg). (**C**) %C-Alb values in animals fed low- or normal-protein diets supplemented with or without urea (67 mg per gram of feed). Bars indicate mean ± SD, *n* = 6 animals per group. *P* values were calculated with an unpaired *t* test.

signaling, and endothelial/monocyte cell adhesion, each of which may exacerbate atherogenesis even further (27, 28, 35). More significantly, when nephrectomized apoE-null mice were given supplemental urea in their drinking water, their rate of atherogenesis increased twofold (12).

We have shown that amino acid concentrations can modify protein carbamylation rates in vitro and in animals. If protein carbamylation is contributing to CVD in ESRD patients, amino acid supplementation may represent a valuable therapeutic approach to improving survival in these patients. Free amino acids are important nucleophiles that may be quantitatively consumed by carbamylation in patients with ESRD (33, 34). N-carbamylation neutralizes amino acids' ability to scavenge oxidants such as hypochlorous acid (48). S-carbamylation also neutralizes the antioxidant properties of thiol amino acids and glutathione (48). Consumptive amino acid carbamylation may thus contribute to oxidative stress. Conversely, treatment of patients with free amino acid scavengers may not only counteract protein carbamylation but also combat the buildup of oxidants in CKD and ESRD.

Here, we have also presented evidence that %C-Alb values correlate significantly with concentrations of urea in the blood. %C-Alb may provide a clearer index of average urea concentrations than isolated blood urea measurements because blood urea can fluctuate 40 to 70% around any given dialysis session. Albumin has a half-life of about 2 weeks in ESRD patients, and carbamylated albumin would therefore represent an index of the time-averaged urea/amino acid balance (49).

It is important to note the limitations of the present study as well as the remaining questions that warrant further investigation: First, our experiments do not test whether circulating carbamylated albumin has any direct pathologic role in CVD or mortality. Recent experiments have shown pathogenic effects by carbamylated albumin ligand on certain tissues (47), but further investigation is required to determine whether a receptor-mediated signaling cascade is involved or whether the effects of carbamylated albumin contribute to uremia-associated sequelae in vivo. Second, our outcome studies focused on 1-year mortality in ESRD patients receiving HD. Further studies are required to determine whether %C-Alb is also a risk factor for patients in earlier stages of CKD. As shown in Fig. 2A, average %C-Alb values are elevated in CKD and ESRD patients compared to non-uremic controls and were largely equivalent in the HD and predialysis CKD cohorts. This indicates that "hypercarbamylation" is present throughout the stages of kidney disease before and after initiation of HD. Further studies are required to determine whether carbamylation is pathogenic at earlier stages of disease and whether predialysis CKD patients may also benefit from amino acid therapy or treatment with pharmacological carbamylation scavengers. Third, although the human observational studies, animal model, and in vitro experiments described here suggest a role for free amino acids in regulating protein carbamylation, they do not demonstrate whether amino acid therapy in human patients will reduce protein carbamylation. Human interventional studies in human ESRD subjects are needed to test whether protein carbamylation (high %C-Alb) decreases in response to free amino acid supplementation or other nucleophilic scavengers and whether these reductions in protein carbamylation are associated with improved patient survival. Furthermore, although our in vitro studies of albumin carbamylation suggest that certain amino acids are more effective scavengers than others, our studies do not address what kinds of scavengers would be most effective in human patients. Differences in nucleophilicity and reactivity to cyanate, differences in pharmacokinetics, and differences in the safety and tolerability of various amino acid scavengers could affect their therapeutic effectiveness. Additional studies are needed to find the optimal combination of amino acids for the prevention of protein carbamylation. Finally, although our results show that high %C-Alb is a significant risk factor for mortality in HD patients, our results cannot be used to interpret %C-Alb measurements in the clinical setting or predict the risk of death in individual ESRD patients. Additional prospective clinical trials to better define

the prognostic value of this test and its ability to predict response to therapy are needed for this blood test to be translated for clinical use.

In summary, our study describes an assay for protein carbamylation (%C-Alb) that was strongly and more reproducibly associated with risk of death than other standard risk factors in two large independent ESRD cohorts. Amino acid deficiencies in ESRD patients may exacerbate protein carbamylation, suggesting an easily modifiable mechanism that contributes to protein carbamylation. Protein carbamylation may thus represent a new modifiable risk factor for mortality in ESRD, providing an attractive target for future therapeutic trials.

MATERIALS AND METHODS

Materials

All chemicals, except when noted, were purchased from Sigma-Aldrich.

Study populations

Frozen specimens from 187 participants of the ArMORR study and 1161 subjects of 4D trial were used for analysis (4, 18, 19, 50). ArMORR subjects' baseline biochemical data and characteristics (table S3) were measured after the 90-day run-in period and just before initiation of the survival study. 4D subjects' baseline characteristics and chemistries (table S4) were collected 1 week before subject randomization. Additional details on ArMORR and 4D study populations can be found in the Supplementary Materials.

Biochemical analyses

Instrumentation. All LC-MS/MS analyses were performed with reversed-phase chromatography coupled to MS/MS. The experiment shown in Fig. 1 was performed on an API 5600 time-of-flight mass spectrometer; experiments shown in table S1 were performed on an API 5000 triple quadrupole mass spectrometer. All other measurements of carbamylated albumin, glycated albumin, carbamylated hemoglobin, and serum amino acids were measured on an API 3200 mass spectrometer. All instruments were purchased from AB Sciex.

Measurement of human serum %C-Alb. Serum samples were mixed with isotopic peptide standards and then digested with Glu-C protease for peptide analysis by LC-MS/MS. Digested serum proteins were injected onto a C18 reversed-phase column, and peptides were separated by gradient elution using conditions described in table S9. The carbamylated and noncarbamylated peptides of interest and their isotopic standards were measured by MRM monitoring using MS/MS mass transitions, and settings are shown in table S9 and quantified according to their area under the curve (AUC) of the eluting peaks (fig. S1). Intra-assay measurement of the 5% carbamylated isotopic peptide standard mix was used to adjust and calibrate assay measurements so that the absolute %C-Alb could be calculated using the following equation: $%C-Alb = 5\% \times$ (carbamylated albumin AUC/noncarbamylated albumin AUC)/(carbamylated isotopic standard AUC/noncarbamylated isotopic standard AUC). The %C-Alb assay demonstrated excellent linearity $(r^2 = 0.9964, y = 0.9826x - 0.0002)$ (51) and coefficient of variation (CV) of 4.2% (fig. S1). Replicate measurements (n = 8) of a random serum sample demonstrated an SD of 0.06% (CV = 4.2%). Details describing specimen processing and additional methods for biochemical analysis of mouse %C-Alb, glycated albumin, carbamylated albumin, and amino acid analysis can be found in the Supplementary Materials.

In vitro experiments

Search for carbamylation sites on human albumin. Purified human albumin (40 g/liter) dissolved in phosphate buffer (pH 7.4) was carbamylated overnight with 1 mM potassium cyanate. Albumin was reduced with dithiothreitol, heat-denatured, and digested with Glu-C protease. Digested peptides were analyzed on an API 5600 mass spectrometer. MS/MS fragmentation spectra were matched against the Swiss-Prot human proteome with Protein Pilot software (AB Sciex).

Relative quantification of carbamylation sites on cyanatetreated and untreated albumin. Serum from an ESRD patient was treated with and without 10 mM potassium cyanate and digested with Glu-C. Percent carbamylation on different lysines within albumin was performed by measuring each lysine-containing albumin peptide and its carbamylated form using mass settings shown in table S1, integrating their AUC values, and calculating the ratios of the carbamylated and noncarbamylated forms.

Carbamylation of amino acid mixtures in vitro. Cyanate was titrated into equimolar mixtures of purified amino acids (250 μ M) in 100 mM phosphate buffer (pH 7.4), and carbamylated amino acids were analyzed by LC-MS/MS.

In vitro inhibition of albumin carbamylation by amino acids. Purified albumin (40 g/liter) was premixed with phosphate buffer (pH 7.4) (100 mM final concentration) and 10 mM of individual amino acids, followed by addition of 0.5 mM cyanate and overnight incubation at 37°C, and then analysis of %C-Alb by LC-MS/MS.

Mouse model of amino acid deficiency and cyanate-induced carbamylation. Two groups of 10-week-old male C57BL/6J mice (n = 6 per group) were given a low-protein diet for 15 days to induce serum amino acid deficiencies (52). Two additional control groups were given feed with normal-protein content. Diets were prepared by Research Diets Inc. and were based on rodent diet AIN-76A. The normal-protein diet contained 20% casein. The low-protein diet contained 2% casein and was made isocaloric to the normal diet by supplementing with sucrose.

Chronic carbamylation by urea. One group of mice on low-protein diet and one group on normal-protein diet also had urea added to their feed (67 mg of urea per gram of feed) throughout the 15-day study.

Acute carbamylation by cyanate. On day 15 of the study, one group of mice on low-protein diet and one group on normal-protein diet were anesthetized with isoflurane, had blood drawn, and were injected intraperitoneally with sodium cyanate to induce carbamylation (100 mg of cyanate per kilogram of body weight) (53). Thirty minutes after injection, animals had blood drawn again followed by euthanasia by carbon dioxide asphyxiation. Animals were anesthetized throughout the procedure. After blood was drawn, samples were allowed to clot and serum was separated, snap-frozen in liquid nitrogen, and frozen at -80° C for later analysis. Samples were analyzed for %C-Alb and free amino acid concentrations. All procedures were performed according to the Institutional Animal Care and Use Committee–approved protocol no. 051-2012/101118 (Animal Welfare Assurance A3153-01).

Statistical analysis

SAS software, version 9.2 (SAS Institute), was used for all analyses. Comparisons between characteristics of subject groups were analyzed with Student's *t* test or Mann-Whitney *U* tests or χ^2 tests where appropriate. The Kaplan-Meier product-limit method was used for visualization of time-to-event variables. Analysis of all-cause mortality was performed with a Cox proportional hazards models. Proportional hazard assumptions were checked for all models. %C-Alb values were transformed to their natural log and analyzed as a continuous variable for HR analysis. The variables included in the final multivariate model were limited to just those that demonstrated statistically significant risks (P < 0.05) during univariate HR analysis. Associations between carbamylated albumin and total amino acids were analyzed with Spearman's rank correlation coefficients, and partial correlations were used to adjust for relevant covariates. Multiple correlations were assessed with multiple linear regression for natural log-transformed outcomes and covariates. Overall significance of comparisons of carbamylated albumin by amino acid tertile was performed with Kruskal-Wallis tests, and significant associations were further explored with Sidak-adjusted, Wilcoxon rank sum tests. Two-tailed P values of <0.05 were considered significant.

SUPPLEMENTARY MATERIALS

www.sciencetranslationalmedicine.org/cgi/content/full/5/175/175ra29/DC1 Materials and Methods

Fig. S1. Development and validation of assay for percent carbamylated albumin.

Fig. S2. Correlations between percent carbamylated albumin, percent carbamylated lysine in serum total protein, and percent carbamylated hemoglobin.

Fig. S3. Dose-dependent carbamylation and depletion of free amino acids in vitro by increasing concentrations of cyanate.

Fig. S4. Correlation between %C-Alb and relative amounts of percent glycated albumin.

Fig. S5. Correlation between %C-Alb and serum albumin concentrations.

Fig. S6. Flow chart describing ArMORR subject acquisition and selection process.

Table S1. Relative quantitation of percent carbamylated lysine within human albumin treated with 10 mM cyanate.

Table S2. Individual %C-Alb values for subjects in groups shown in Fig. 2.

- Table S3. Characteristics of ArMORR study population by mortality status.
- Table S4. Characteristics of 4D study population by mortality status.
- Table S5. ArMORR and 4D study HR estimates for mortality by tertile of %C-Alb.

Table S6. Characteristics of ArMORR study cases and 12-month survivors used in this study along with their original matched cohorts.

Table S7. Characteristics of stage 3 and 4 CKD subjects.

Table S8. Characteristics of non-uremic subjects.

Table S9. HPLC and MS settings.

REFERENCES AND NOTES

- G. Eknoyan, N. Lameire, R. Barsoum, K. U. Eckardt, A. Levin, N. Levin, F. Locatelli, A. MacLeod, R. Vanholder, R. Walker, H. Wang, The burden of kidney disease: Improving global outcomes. *Kidney Int.* 66, 1310–1314 (2004).
- R. N. Foley, P. S. Parfrey, M. J. Sarnak, Clinical epidemiology of cardiovascular disease in chronic renal disease. Am. J. Kidney Dis. 32, S112–S119 (1998).
- United States Renal Data System, 2009 Annual Data Report: Atlas of Chronic Kidney Disease and End-Stage Renal Disease (National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases, Bethesda, MD, 2009).
- C. Wanner, V. Krane, W. März, M. Olschewski, J. F. Mann, G. Ruf, E. Ritz; German Diabetes and Dialysis Study Investigators, Atorvastatin in patients with type 2 diabetes mellitus undergoing hemodialysis. *N. Engl. J. Med.* **353**, 238–248 (2005).
- B. C. Fellström, A. G. Jardine, R. E. Schmieder, H. Holdaas, K. Bannister, J. Beutler, D. W. Chae, A. Chevaile, S. M. Cobbe, C. Grönhagen-Riska, J. J. De Lima, R. Lins, G. Mayer, A. W. McMahon, H. H. Parving, G. Remuzzi, O. Samuelsson, S. Sonkodi, D. Sci, G. Süleymanlar, D. Tsakiris, V. Tesar, V. Todorov, A. Wiecek, R. P. Wuthrich, M. Gottlow, E. Johnsson, F. Zannad; AURORA Study Group, Rosuvastatin and cardiovascular events in patients undergoing hemodialysis. *N. Engl. J. Med.* **360**, 1395–1407 (2009).
- S. A. Karumanchi, R. Thadhani, Kidney complications: Why don't statins always work? *Nat. Med.* 16, 38–40 (2010).
- W. J. Johnson, W. W. Hagge, R. D. Wagoner, R. P. Dinapoli, J. W. Rosevear, Effects of urea loading in patients with far-advanced renal failure. *Mayo Clin. Proc.* 47, 21–29 (1972).
- G. M. Chertow, W. F. Owen, J. M. Lazarus, N. L. Lew, E. G. Lowrie, Exploring the reverse J-shaped curve between urea reduction ratio and mortality. *Kidney Int.* 56, 1872–1878 (1999).

- M. Stosovic, M. Stanojevic, S. Simic-Ogrizovic, D. Jovanovic, L. Djukanovic, Relation between serum urea and mortality of hemodialysis patients. *Ren. Fail.* **31**, 335–340 (2009).
- G. Eknoyan, G. J. Beck, A. K. Cheung, J. T. Daugirdas, T. Greene, J. W. Kusek, M. Allon, J. Bailey, J. A. Delmez, T. A. Depner, J. T. Dwyer, A. S. Levey, N. W. Levin, E. Milford, D. B. Ornt, M. V. Rocco, G. Schulman, S. J. Schwab, B. P. Teehan, R. Toto; Hemodialysis (HEMO) Study Group, Effect of dialysis dose and membrane flux in maintenance hemodialysis. *N. Engl. J. Med.* 347, 2010–2019 (2002).
- Z. Wang, S. J. Nicholls, E. R. Rodriguez, O. Kummu, S. Hörkkö, J. Barnard, W. F. Reynolds, E. J. Topol, J. A. DiDonato, S. L. Hazen, Protein carbamylation links inflammation, smoking, uremia and atherogenesis. *Nat. Med.* **13**, 1176–1184 (2007).
- E. O. Apostolov, D. Ray, A. V. Savenka, S. V. Shah, A. G. Basnakian, Chronic uremia stimulates LDL carbamylation and atherosclerosis. J. Am. Soc. Nephrol. 21, 1852–1857 (2010).
- R. Flückiger, W. Harmon, W. Meier, S. Loo, K. H. Gabbay, Hemoglobin carbamylation in uremia. *N. Engl. J. Med.* 304, 823–827 (1981).
- M. Oimomi, K. Ishikawa, T. Kawasaki, S. Kubota, Y. Yoshimura, S. Baba, Plasma carbamylated protein in renal failure. N. Engl. J. Med. 308, 655–656 (1983).
- G. R. Stark, W. H. Stein, S. Moore, Reactions of the cyanate present in aqueous urea with amino acids and proteins. J. Biol. Chem. 235, 3177–3181 (1960).
- D. M. Nathan, J. Kuenen, R. Borg, H. Zheng, D. Schoenfeld, R. J. Heine; A1c-Derived Average Glucose Study Group, Translating the A1C assay into estimated average glucose values. *Diabetes Care* **31**, 1473–1478 (2008).
- A. Frolov, R. Hoffmann, Identification and relative quantification of specific glycation sites in human serum albumin. *Anal. Bioanal. Chem.* 397, 2349–2356 (2010).
- S. M. Brunelli, R. I. Thadhani, K. E. Lynch, E. D. Ankers, M. M. Joffe, R. Boston, Y. Chang, H. I. Feldman, Association between long-term blood pressure variability and mortality among incident hemodialysis patients. *Am. J. Kidney Dis.* 52, 716–726 (2008).
- O. M. Gutiérrez, M. Mannstadt, T. Isakova, J. A. Rauh-Hain, H. Tamez, A. Shah, K. Smith, H. Lee, R. Thadhani, H. Jüppner, M. Wolf, Fibroblast growth factor 23 and mortality among patients undergoing hemodialysis. *N. Engl. J. Med.* **359**, 584–592 (2008).
- 20. B. Rosner, Fundamentals of Biostatistics (Duxbury Press, Belmont, CA, ed. 6, 2006).
- J. P. Klein, M. L. Moeschberger, Survival Analysis: Techniques for Censored and Truncated Data (Springer, New York, ed. 2, 2003).
- T. Depner, G. Beck, J. Daugirdas, J. Kusek, G. Eknoyan, Lessons from the Hemodialysis (HEMO) Study: An improved measure of the actual hemodialysis dose. *Am. J. Kidney Dis.* 33, 142–149 (1999).
- C. Wanner, V. Krane, W. März, M. Olschewski, H. G. Asmus, W. Krämer, K. W. Kühn, H. Kütemeyer, J. F. Mann, G. Ruf, E. Ritz; Deutsche Diabetes-Dialyse-Studie (4D) Study Group, Randomized controlled trial on the efficacy and safety of atorvastatin in patients with type 2 diabetes on hemodialysis (4D study): Demographic and baseline characteristics. *Kidney Blood Press. Res.* 27, 259–266 (2004).
- B. Yang, L. Bankir, Urea and urine concentrating ability: New insights from studies in mice. Am. J. Physiol. Renal Physiol. 288, F881–F896 (2005).
- P. Goldwasser, N. Mittman, A. Antignani, D. Burrell, M. A. Michel, J. Collier, M. M. Avram, Predictors of mortality in hemodialysis patients. J. Am. Soc. Nephrol. 3, 1613–1622 (1993).
- S. Malgorzewicz, A. Debska-Slizien, B. Rutkowski, W. Lysiak-Szydlowska, Serum concentration of amino acids versus nutritional status in hemodialysis patients. J. Ren. Nutr. 18, 239–247 (2008).
- E. O. Apostolov, S. V. Shah, E. Ok, A. G. Basnakian, Carbamylated low-density lipoprotein induces monocyte adhesion to endothelial cells through intercellular adhesion molecule-1 and vascular cell adhesion molecule-1. *Arterioscler. Thromb. Vasc. Biol.* 27, 826–832 (2007).
- E. O. Apostolov, S. V. Shah, D. Ray, A. G. Basnakian, Scavenger receptors of endothelial cells mediate the uptake and cellular proatherogenic effects of carbamylated LDL. *Arterioscler*. *Thromb. Vasc. Biol.* 29, 1622–1630 (2009).
- G. Asci, A. Basci, S. V. Shah, A. Basnakian, H. Toz, M. Ozkahya, S. Duman, E. Ok, Carbamylated low-density lipoprotein induces proliferation and increases adhesion molecule expression of human coronary artery smooth muscle cells. *Nephrology* 13, 480–486 (2008).
- C. M. Balion, T. F. Draisey, R. J. Thibert, Carbamylated hemoglobin and carbamylated plasma protein in hemodialyzed patients. *Kidney Int.* 53, 488–495 (1998).
- J. Carracedo, A. Merino, C. Briceño, S. Soriano, P. Buendía, L. Calleros, M. Rodriguez, A. Martín-Malo, P. Aljama, R. Ramírez, Carbamylated low-density lipoprotein induces oxidative stress and accelerated senescence in human endothelial progenitor cells. *FASEB J.* 25, 1314–1322 (2011).
- S. Jaisson, C. Delevallée-Forte, F. Touré, P. Rieu, R. Garnotel, P. Gillery, Carbamylated albumin is a potent inhibitor of polymorphonuclear neutrophil respiratory burst. *FEBS Lett.* 581, 1509–1513 (2007).
- L. M. Kraus, M. R. Jones, A. P. Kraus Jr., Essential carbamoyl-amino acids formed in vivo in patients with end-stage renal disease managed by continuous ambulatory peritoneal dialysis: Isolation, identification, and quantitation. J. Lab. Clin. Med. 131, 425–431 (1998).
- L. M. Kraus, A. P. Kraus Jr., Carbamoylation of amino acids and proteins in uremia. *Kidney* Int. Suppl. 78, S102–S107 (2001).
- E. Ok, A. G. Basnakian, E. O. Apostolov, Y. M. Barri, S. V. Shah, Carbamylated low-density lipoprotein induces death of endothelial cells: A link to atherosclerosis in patients with kidney disease. *Kidney Int.* 68, 173–178 (2005).

- G. R. Stark, On the reversible reaction of cyanate with sulfhydryl groups and the determination of NH₂-terminal cysteine and cystine in proteins. J. Biol. Chem. 239, 1411–1414 (1964).
- G. R. Stark, Reactions of cyanate with functional groups of proteins. 3. Reactions with amino and carboxyl groups. *Biochemistry* 4, 1030–1036 (1965).
- G. R. Stark, Reactions of cyanate with functional groups of proteins. IV. Inertness of aliphatic hydroxyl groups. Formation of carbamyl- and acylhydantoins. *Biochemistry* 4, 2363–2367 (1965).
- G. R. Stark, Reactions of cyanate with functional groups of proteins. II. Formation, decomposition, and properties of *N*-carbamylimidazole. *Biochemistry* 4, 588–595 (1965).
- G. R. Stark, D. G. Smyth, The use of cyanate for the determination of NH₂-terminal residues in proteins. J. Biol. Chem. 238, 214–226 (1963).
- S. Jaisson, C. Pietrement, P. Gillery, Carbamylation-derived products: Bioactive compounds and potential biomarkers in chronic renal failure and atherosclerosis. *Clin. Chem.* 57, 1499–1505 (2011).
- R. Paniagua, D. Amato, E. Vonesh, R. Correa-Rotter, A. Ramos, J. Moran, S. Mujais; Mexican Nephrology Collaborative Study Group, Effects of increased peritoneal clearances on mortality rates in peritoneal dialysis: ADEMEX, a prospective, randomized, controlled trial. J. Am. Soc. Nephrol. 13, 1307–1320 (2002).
- L. Elangovan, C. S. Shinaberger, J. A. Kraut, J. H. Shinaberger, HEMO equilibrated Kt/V goals are difficult to achieve in large male patients. ASAIO J. 47, 235–239 (2001).
- J. C. Ayus, M. R. Mizani, S. G. Achinger, R. Thadhani, A. S. Go, S. Lee, Effects of short daily versus conventional hemodialysis on left ventricular hypertrophy and inflammatory markers: A prospective, controlled study. J. Am. Soc. Nephrol. 16, 2778–2788 (2005).
- R. P. Pauly, Nocturnal home hemodialysis and short daily hemodialysis compared with kidney transplantation: Emerging data in a new era. Adv. Chronic Kidney Dis. 16, 169–172 (2009).
- R. A. Koeth, K. Kalantar-Zadeh, Z. Wang, S. L. Hazen, Abstract 1496: Protein bound homocitrulline independently predicts mortality risks in maintenance hemodialysis patients. *Circulation* 118, S_337 (2008).
- M. L. Gross, G. Piecha, A. Bierhaus, W. Hanke, T. Henle, P. Schirmacher, E. Ritz, Glycated and carbamylated albumin are more "nephrotoxic" than unmodified albumin in the amphibian kidney. *Am. J. Physiol. Renal Physiol.* **301**, F476–F485 (2011).
- S. M. Schreier, M. Hollaus, M. Hermann, L. Jirovetz, M. Exner, S. Kapiotis, B. M. Gmeiner, H. Laggner, Carbamoylated free amino acids in uremia: HOCI generates volatile protein modifying and cytotoxic oxidant species from N-carbamoyl-threonine but not threonine. *Biochimie* **94**, 2441–2447 (2012).
- J. C. Fish, A. R. Remmers Jr., J. D. Lindley, H. E. Sarles, Albumin kinetics and nutritional rehabilitation in the unattended home-dialysis patient. *N. Engl. J. Med.* 287, 478–481 (1972).
- D. J. Friedman, M. Afkarian, H. Tamez, I. Bhan, T. Isakova, M. Wolf, E. Ankers, J. Ye, M. Tonelli, C. Zoccali, M. Kuro-o, O. Moe, S. A. Karumanchi, R. Thadhani, Klotho variants and chronic hemodialysis mortality. *J. Bone Miner. Res.* 24, 1847–1855 (2009).

- S. Lin, T. A. Shaler, C. H. Becker, Quantification of intermediate-abundance proteins in serum by multiple reaction monitoring mass spectrometry in a single-quadrupole ion trap. *Anal. Chem.* **78**, 5762–5767 (2006).
- J. E. Antflick, G. B. Baker, D. R. Hampson, The effects of a low protein diet on amino acids and enzymes in the serine synthesis pathway in mice. *Amino Acids* 39, 145–153 (2010).
- J. Tor-Agbidye, V. S. Palmer, P. S. Spencer, A. M. Craig, L. L. Blythe, M. I. Sabri, Sodium cyanate alters glutathione homeostasis in rodent brain: Relationship to neurodegenerative diseases in protein-deficient malnourished populations in Africa. *Brain Res.* 820, 12–19 (1999).
- I. V. Shilov, S. L. Seymour, A. A. Patel, A. Loboda, W. H. Tang, S. P. Keating, C. L. Hunter, L. M. Nuwaysir, D. A. Schaeffer, The Paragon Algorithm, a next generation search engine that uses sequence temperature values and feature probabilities to identify peptides from tandem mass spectra. *Mol. Cell. Proteomics* 6, 1638–1655 (2007).

Funding: Supported by the Beth Israel Deaconess Medical Center Department of Pathology (to A.H.B.); the Howard Hughes Medical Institute (to S.A.K.); K24 DK094872 (to R.T.); and the University Hospital Wüerzberg "Gundausstattung" grant program (to C.D. and C.W.). Author contributions: A.H.B. and S.A.K. contributed to the study conception and design. A.H.B. performed assay development, experimental design, biochemical analyses, data interpretation, and primary preparation of the manuscript. S.A.K. provided assistance with experimental design, data interpretation, and costs associated with study. R.T. provided ArMORR study specimens. R.T. and J.W. performed data analysis associated with the ArMORR study. C.W. and C.D. provided specimens and data analysis associated with the 4D study. W.R., A.H.B., and S.A.K. performed animal experiments, D.J.F. and J.D. provided specimens and data analysis for CKD study. T.H. and R.B. assisted with biochemical analyses. H.S. performed high mass resolution LC-MS/MS analysis to confirm carbamylation sites on human albumin, C.D., J.W., R.B., S.K., S.M.P., J.D., D.J.F., R.T., and S.A.K. provided assistance with review and preparation of manuscript. Competing interests: Provisional applications for U.S. and International patents related to the contents of this manuscript have been filed by A.H.B., S.A.K., R.T., and their affiliated institutions. R.T. has served as a consultant to Fresenius Medical Care.

Submitted 24 October 2012 Accepted 11 February 2013 Published 6 March 2013 10.1126/scitranslmed.3005218

Citation: A. H. Berg, C. Drechsler, J. Wenger, R. Buccafusca, T. Hod, S. Kalim, W. Ramma, S. M. Parikh, H. Steen, D. J. Friedman, J. Danziger, C. Wanner, R. Thadhani, S. A. Karumanchi, Carbamylation of serum albumin as a risk factor for mortality in patients with kidney failure. *Sci. Transl. Med.* **5**, 175ra29 (2013).





Carbamylation of Serum Albumin as a Risk Factor for Mortality in Patients with Kidney Failure Anders H. Berg, Christiane Drechsler, Julia Wenger, Roberto Buccafusca, Tammy Hod, Sahir Kalim, Wenda Ramma, Samir M. Parikh, Hanno Steen, David J. Friedman, John Danziger, Christoph Wanner, Ravi Thadhani and S. Ananth Karumanchi (March 6, 2013)

Science Translational Medicine **5** (175), 175ra29. [doi: 10.1126/scitranslmed.3005218]

Editor's Summary

Counteracting Carbamylation: A Possible Route to Treating Complications of Kidney Failure

Like a canary in a coal mine, too much glycated hemoglobin in the blood is a warning sign that a diabetic patients' glucose is out of control. Carbamyl groups on another ubiquitous blood protein, albumin, may sound a similar alarm for blood urea, a consequence of failing kidneys. Carbamylation may be harmful in its own right because it has been linked to atherosclerosis and other diseases.

By using mass spectroscopy, the authors devised a highly accurate assay for measuring carbamylation of the lysine at position 549 of human albumin. With this assay, they found that in two groups of patients with end-stage renal disease and elevated blood urea, the amount of carbamylated albumin correlated with urea concentrations. Albumin was more carbamylated in patients with end-stage renal disease who died within a year than in those who lived longer.

Although knowing the prognosis of patients with kidney failure would let us prioritize transplants for the most needy, it would be even better if we could prevent the complications of kidney failure altogether. The authors have gathered data from biochemical experiments and from mice that suggest that protein carbamylation and its associated pathology might be prevented by boosting amino acid concentrations (which tend to be low in these patients) in the blood through diet or other means. Amino acids can compete with protein side chains during carbamylation, potentially interfering with the reaction and its harmful consequences. Thus, amino acid replacement might help patients with kidney failure.

The following resources related to this article are available online at http://stm.sciencemag.org. This information is current as of January 21, 2016.

Article Tools	Visit the online version of this article to access the personalization and article tools: http://stm.sciencemag.org/content/5/175/175ra29
Supplemental	"Supplementary Materials"
Materials	http://stm.sciencemag.org/content/suppl/2013/03/04/5.175.175ra29.DC1
Science Translational M	<i>Medicine</i> (print ISSN 1946-6234; online ISSN 1946-6242) is published
weekly, except the last y	week in December, by the American Association for the Advancement of
Science, 1200 New Yor	k Avenue, NW, Washington, DC 20005, Copyright 2016 by the American

Association for the Advancement of Science; all rights reserved. The title Science Translational

Medicine is a registered trademark of AAAS.

Related Content	The editors suggest related resources on <i>Science</i> 's sites: http://stm.sciencemag.org/content/scitransmed/1/5/5ra13.full http://stm.sciencemag.org/content/scitransmed/5/172/172ra22.full http://stm.sciencemag.org/content/scitransmed/3/85/85ra46.full http://stm.sciencemag.org/content/scitransmed/5/198/198ra106.full http://stm.sciencemag.org/content/scitransmed/4/121/121ra18.full http://stm.sciencemag.org/content/scitransmed/7/316/316ra193.full
Doumiasiona	Obtain information about reproducing this articles

Permissions Obtain information about reproducing this article: http://www.sciencemag.org/about/permissions.dtl

Science Translational Medicine (print ISSN 1946-6234; online ISSN 1946-6242) is published weekly, except the last week in December, by the American Association for the Advancement of Science, 1200 New York Avenue, NW, Washington, DC 20005. Copyright 2016 by the American Association for the Advancement of Science; all rights reserved. The title *Science Translational Medicine* is a registered trademark of AAAS.